

## Acetate Incorporation Assay - FAS Activity Assay

This protocol was adapted from: Pizer, E. S., Wood, F. D., Pasternack, G. R., and Kuhajda, F. P. (1996) Cancer Res. 1996, 745–751.

### We've performed this assay on the following cell types:

- Dissociated Primary Cortical Neurons
  - Plate at  $0.5 - 1.0 \times 10^6$  cells/well of a 24 well plate
  - Cultures are used 7-10 days after plating.
- Dissociated Primary Hypothalamic Neurons
  - Plated at  $0.4 - 0.6 \times 10^6$  cells/well of a 24 well plate
  - Cultures are used 7-10 days after plating.

### Drug/Compound treatment:

- Cultures are pre-treated with the compound/drug of interest for 2 hrs @ 37°C.

### Labeling:

- The appropriate amount of  $^3\text{H}$ -acetic acid to use should be determined by performing a titration to determine a concentration that falls within the linear range of the curve. For our neuronal cultures we use a final concentration of 100  $\mu\text{M}$ .
  - Add the appropriate concentration of  $^3\text{H}$ -acetic acid to each well and incubate cultures for an additional 2 hrs @ 37°C.
    - Acetic Acid, Sodium Salt [ $^3\text{H}$ ]; 10.0 mCi/ml; 75-150 mCi/mmol; NEN Life Science Prod. Inc; Cat #NET-003.
  - Following the desired labeling period discard the radioactive media and wash 1X with 1 ml of PBS.

### Extraction:

- Add 900  $\mu\text{l}$  of 2:1 chloroform/methanol to each well. Add 600  $\mu\text{l}$  of 4 mM  $\text{MgCl}_2$  one well at a time. Triturate **2-3 times** (until homogenous) and transfer to eppi. tube.
- Vortex and spin for 1 min at high speed. Discard aqueous phase (upper) leaving a little aqueous phase to minimize error.
- Repeat extraction 3 more times with varying amounts of chloroform/methanol and  $\text{MgCl}_2$ :
  - 650  $\mu\text{l}$  chloroform/methanol and 450  $\mu\text{l}$  4 mM and  $\text{MgCl}_2$
  - 400  $\mu\text{l}$  chloroform/methanol and 200  $\mu\text{l}$  4 mM and  $\text{MgCl}_2$
  - 300  $\mu\text{l}$  chloroform/methanol and 100  $\mu\text{l}$  4 mM and  $\text{MgCl}_2$

***\*\*It is important to remove all of the aqueous phase after the last extraction.***

- Transfer the organic phase to a scintillation vial and dry under  $\text{N}_2$  for approx 5-10 min at 50-70 °C.
- Resuspend in 500  $\mu\text{l}$  of 100% Ethanol then add **4 mls** of scintillation fluid and count the entire sample OR resuspend directly in scintillation fluid.