

Hackam Lab IHC for Whole Mount

REAGENTS

1. **4% PFA:** Obtain 16% PFA, place into 50cc tube and add PBS to 40 mL.
2. **PBS-PFA-0.1% Triton x 100:** Add Triton x 100 (40 μ L) into 40mL of 4% PFA.
3. **10% & 1% Normal Donkey Serum (NDS) in PBST**

METHOD

Day 1

1. Remove the media from chamber slides
2. Wash the slides with PBS (x 1).
3. Place 300 μ L of PBS-PFA-0.1% Triton X 100 into each well for 1 hr at RT.
4. Wash with PBS for **15 min** at RT (x 4).
5. Block with 10% NDS in PBST for **45 min** at RT.
6. Incubate with primary antibody in 1% NDS/PBST in 4 degree fridge on rotator **overnight**. Dilution condition of most primary antibodies is **1:250**.

Day 2

7. Wash with PBST on rotator in 4 degree for 4-5 hours changing PBST at least (x 5).
8. Incubate with secondary antibody in 1% NDS/PBST in 4 degree fridge on rotator **overnight**. Dilution condition of secondary antibody is **1:1000**.

Day 3

9. Add Dapi directly to the secondary antibody (add double the amount already within the well) for 15 min.
 10. Wash with PBST on rotator in 4 degree for 10 minutes (x 6).
 11. Remove the wells with the razor blade and tacking.
 12. Place mounting gel without bubbles on each well.
- Place coverslip and place into dark, dry shelf.